

Isotopic record of penguin diet



Neandertal DNA damage

The protein sequence flow network

Brain high-field phase MRI

Lignin formation in pine

Abrupt recent shift in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values in Adélie penguin eggshell in Antarctica

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Stable isotope values of carbon ($\delta^{13}\text{C}$) and nitrogen ($\delta^{15}\text{N}$) in blood, feathers, eggshell, and bone have been used in seabird studies since the 1980s, providing a valuable source of information on diet, foraging patterns, and migratory behavior in these birds. These techniques can also be applied to fossil material when preservation of bone and other tissues is sufficient. Excavations of abandoned Adélie penguin (*Pygoscelis adeliae*) colonies in Antarctica often provide well preserved remains of bone, feathers, and eggshell dating from hundreds to thousands of years B.P. Herein we present an $\approx 38,000$ -year time series of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of Adélie penguin eggshell from abandoned colonies located in three major regions of Antarctica. Results indicate an abrupt shift to lower-trophic prey in penguin diets within the past ≈ 200 years. We posit that penguins only recently began to rely on krill as a major portion of their diet, in conjunction with the removal of baleen whales and krill-eating seals during the historic whaling era. Our results support the “krill surplus” hypothesis that predicts excess krill availability in the Southern Ocean after this period of exploitation.

abandoned colonies | stable isotopes | krill surplus | dietary shift | historic whaling

Abandoned colonies of Adélie penguins (*Pygoscelis adeliae*) have been located in many coastal, ice-free regions of Antarctica. When excavated, these sites often provide abundant well preserved remains of penguin tissue (bone, feathers, dried skin, and eggshell) and hard parts from prey remains (fish bone, otoliths, and squid beaks) in guano. Radiocarbon dates on these tissues range from hundreds to tens of thousands of years old, providing considerable information on the occupation history and population movements of Adélie penguins from the late Pleistocene through Holocene in the Antarctic Peninsula (1–3), Ross Sea (4–6), and East Antarctica (7). Although these studies have been expanded in recent years to include analysis of ancient DNA (8, 9), until now stable isotope analyses have not been applied except for characterization of living specimens.

Since the 1980s, carbon ($\delta^{13}\text{C}$) and nitrogen ($\delta^{15}\text{N}$) stable isotope values of seabird tissues have been used to address questions on diet and foraging behavior (10–14). These studies are particularly useful when traditional diet studies (observations of chick feedings and stomach lavage) are difficult to carry out on far-ranging species such as many seabirds. For penguins in Antarctica stable isotope analyses allow assessment of seasonal differences in diet among widely dispersed populations.

Modern Adélie penguins are known to feed primarily on krill (*Euphausia superba*) during the chick-rearing period in the Antarctic Peninsula, although Antarctic silverfish (*Pleurogramma antarcticum*) and squid (*Psychroteuthis antarctica*) can also be important (15). However, because most dietary studies have been conducted during the chick-rearing period, little is known of winter diet (16). Isotope ratios stored in seabird tissues can provide information on diet during brief to long periods, depending on the tissue used, and can provide important new information on diet outside the breeding season. Moreover, fossil eggshell can provide information on dietary shifts with climate change over millennia (17).

We determined $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and $\delta^{18}\text{O}$ values of modern and fossil eggshell recovered from eight active and 28 abandoned colonies of Adélie penguins in Antarctica to determine whether dietary shifts through time could be discerned and compared with environmental and climate change documented in the geological record (ice cores and marine sediments). We chose eggshell for this analysis because it is almost always well preserved in ornithogenic soils (bird-formed soils characteristic of abandoned penguin colonies), whether these soils are hundreds or tens of thousands of years old, whereas other tissues (bone and feathers) are more variably preserved. Because the soils of Antarctica are particularly cold and dry, diagenesis in fossil specimens is unlikely and the original eggshell isotope values should be well preserved. Stable isotope values of eggshell can provide valuable information on the diet of breeding females during a critical stage in the breeding cycle [see Schaffner and Swart (18) for a review of this topic]. Eggshell formation, when initiated, is completed within a 24-h period (19), thus recording the bird's most recent dietary signal. Most female Adélie penguins begin laying eggs in October/November each year (15). Consequently, analyses of eggshells will characterize penguin diet during a brief interval in the annual cycle. To characterize any regional differences in penguin diet, we also examined samples of eggshell from abandoned and active colonies from three major regions of Antarctica: the Ross Sea, East Antarctica, and the Antarctic Peninsula. Samples were collected over several field seasons between 1999 and 2004. All sites were excavated in 5-cm arbitrary levels to the lower boundary of ornithogenic sediments as recognized by a distinct change in soil texture and color [see Emslie *et al.* (5) for excavation methods].

Results and Discussion

More than 220 fossil eggshell samples ranging in age from ≈ 100 to 38,000 years B.P. [reported in either calendar years (cal yr) or ^{14}C yr] and 57 modern samples from the three regions were analyzed for stable carbon, nitrogen, and oxygen isotopes [supporting information (SI) Tables 2 and 3]. Of the fossil samples, 21 fragments were large enough to divide into two pieces, one for radiocarbon dating and one for stable isotope analysis. In this manner it was possible to obtain an absolute date for some of the isotope results. Additional eggshell fragments from the same stratigraphic level as the dated fragments were included in the analysis and assumed to be the same age. Other samples were taken from stratigraphic levels that were dated by using bone found in those levels. In 2004 we sampled 20 historic Adélie

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Abbreviation: cal yr, calendar year; VPDB, Vienna Pee Dee Belemnite.

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Pretoria/Beta Analytic calibration program at Beta Analytic (Coral Gables, FL). These calibrations provide a minimum and maximum age for each sample in cal yr B.P. with a 95% accuracy that the true date of the sample falls within this range. Dates older than 26,000 years could not be calibrated and are reported in ^{14}C yr B.P. To simplify plotting of all calibrated dates with stable isotope data we used the midpoint of each calibrated range in these plots. For samples from undated stratigraphic levels we used the midpoint of the average ages of levels above and below the undated layer.

Selected samples from East Antarctica and the Ross Sea were dated by splitting the eggshell in half and using one part for radiocarbon analysis, by accelerator mass spectrometer, and the other for stable isotope analysis. This method ensured an accurate date on these stable isotope values. Additional isotope samples were dated by association with dated material from the same stratigraphic level. This method assumes that no contamination or mixing of sediment has occurred since the ornithogenic (bird-formed) layers formed. We believe this is a reasonable assumption because multiple dates from stratigraphic levels at the same site, including those on small eggshell fragments and squid beaks, demonstrate that little or no mixing has occurred (2).

Isotope Analyses of Eggshell. Penguin eggshells were prepared for carbonate analyses by crushing with a quartz mortar and pestle until the shells were ground to a fine powder. Stable isotope values were obtained by using a Kiel-III carbonate preparation device (ThermoFinnegan, San Jose, CA) directly coupled to a ThermoFinnegan MAT 253 stable isotope gas ratio mass spectrometer. Eggshell was reacted at 70°C with four drops of 103% anhydrous phosphoric acid for 5 minutes. Isotope values were corrected for acid fractionation and ^{17}O contribution and reported in per mil notation relative to the VPDB standard. The precision and the calibration of data were monitored through

daily analysis of NBS-18 and NBS-19 standards. Precision was $\pm 0.05\%$ for carbon and $\pm 0.07\%$ for oxygen isotope values.

Isotope values of organic components of penguin eggshells were obtained by removal of carbonate within the eggshell matrix by dissolution with ≈ 1.5 ml of a 10% hydrochloric acid solution. Eggshells were rinsed with deionized H_2O and centrifuged repeatedly six times until sample pH was neutral and then freeze-dried. Some authors have observed a minor nitrogen isotope effect related to use of excessive amounts of acid in carbonate-free samples (33). This potential effect is limited in our study both by the buffering capacity of dissolving eggshell calcite and by using only the amount of weak HCl required to dissolve the carbonate. Approximately 0.5 mg of the acid-insoluble fraction was loaded into tin cups and analyzed for carbon and nitrogen isotopes on a ThermoFinnegan EA elemental analyzer via a Conflo-III (continuous flow interface) open-split and processed via continuous flow on a ThermoFinnegan Delta Plus XL stable isotope gas ratio mass spectrometer. Isotope ratios were normalized to VPDB for $\delta^{13}\text{C}$ and AIR for $\delta^{15}\text{N}$ using International Atomic Energy Agency (IAEA) NO-3, IAEA CH-6, and internal standards. Precision was better than $\pm 0.2\%$ for both carbon and nitrogen isotope values. Both of these methods were successfully applied in a preliminary study of fossil and modern penguin eggshell from Antarctica in 2003.

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1. Tatur A, Myrcha A, Niegodzisz J (1997) *Polar Biol* 17:405–417.
2. Emslie SD, Fraser W, Smith RC, Walker W (1998) *Antarct Sci* 10:257–268.
3. Emslie SD (2001) *Antarct Sci* 13:289–295.
4. Baroni C, Orombelli G (1994) *Geology* 22:23–26.
5. Emslie SD, Berkman PA, Ainley DG, Coats L, Polito M (2003) *Mar Ecol Prog Ser* 262:19–25.
6. Emslie SD, Coats L, Licht K (2007) *Geology* 35:61–64.
7. Emslie SD, Woehler EJ (2005) *Antarct Sci* 17:57–66.
8. Lambert DM, Ritchie PA, Millar CD, Holland B, Drummond AJ, Baroni C (2002) *Science* 295:2270–2273.
9. Shepherd LD, Millar CD, Ballard G, Ainley DG, Wilson PR, Haynes GD, Baroni C, Lambert DM (2005) *Proc Natl Acad Sci USA* 102:16717–16722.
10. Hobson DA (1987) *J Can Zool* 65:1210–1213.
11. Hobson KA (1995) *Condor* 97:752–762.
12. Hobson KA, Piatt JF, Pitocchelli J (1994) *J Anim Ecol* 63:786–798.
13. Hodum PJ, Hobson KA (2000) *Mar Ecol Prog Ser* 198:273–281.
14. Forero MG, Hobson KA, Bortolotti GR, Donazar JA, Bertellotti M, Blanco G (2002) *Mar Ecol Prog Ser* 234:289–299.
15. Williams TD (1995) *The Penguins* (Oxford Univ Press, Oxford).
16. Ainley DG (2002) *The Adélie Penguin* (Columbia Univ Press, New York).
17. Johnson BJ, Miller GH, Fogel ML, Magee JW, Gagan MK, Chivas AR (1999) *Science* 284:1150–1152.
18. Schaffner FC, Swart PK (1991) *Bull Mar Sci* 48:23–38.
19. Astheimer LB, Grau CR (1985) *Condor* 87:256–267.
20. Von Schirnding Y, van der Merwe NJ, Vogel JC (1982) *Archaeometry* 24:3–20.
21. Laws RM (1985) *Am Sci* 73:26–40.
22. Croxall JP (1992) *Philos Trans R Soc London B* 338:319–328.
23. Ellis R (1991) *Men and Whales* (Knopf, New York).
24. Fraser WR, Trivelpiece WZ, Ainley DG, Trivelpiece SG (1992) *Polar Biol* 11:525–531.
25. Woehler E (1993) *The Distribution and Abundance of Antarctic and Subantarctic Penguins* (Scientific Committee on Antarctic Res, Cambridge, UK).
26. McGoldrick K, Marris E (2006) *Nature* 444:978–979.
27. Gordon JE, Harkness DD (1992) *Q Sci Rev* 11:697–708.
28. Stuiver M, Reimer PJ, Braziunas TF (1998) *Radiocarbon* 40:1127–1151.
29. Emslie SD, McDaniel J (2002) *Polar Biol* 25:222–229.
30. Emslie SD, Ritchie P, Lambert D (2003) *Antarct Res Ser* 79:171–180.
31. Stuiver M, Reimer PJ (1993) *Radiocarbon* 35:215–230.
32. Hughen KA, Baillie MGL, Bard E, Bayliss A, Beck JW, Blackwell PG, Buck CE, Burr GS, Cutler KB, Damon PE, et al. (2004) *Radiocarbon* 46:1059–1086.
33. Jacob U, Mintenbeck K, Brey T, Knust R, Beyer K (2005) *Mar Ecol Prog Ser* 287:251–253.