

Environ Indicator Sensitivity to Flux Uncertainty in a Phosphorus Model of Lake Sidney Lanier, USA

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Abstract

Effective environmental impact assessment and management requires improved understanding of the organization and transformation of ecosystems in which independent agents are linked through an intricate network of energy, matter, and informational interactions. While advances have been made, we still lack a complete understanding of the processes that create, constrain, and sustain ecosystems. Network Environ Analysis (NEA) provides one approach for building novel ecosystem insights, but it is model dependent. As ecological modeling is an imprecise art, often complicated by inadequate empirical data, the utility of NEA may be limited by model uncertainty. Here, we investigate the sensitivity of NEA indicators of ecosystem growth and development to flow and storage uncertainty in a phosphorus model of Lake Sidney Lanier, USA. The indicators are total system throughflow (*TST*), total system storage (*TSS*), total boundary input (*Boundary*), Finn cycling index (*FCI*), ratio of indirect-to-direct flows (*Indirect/Direct*), indirect flow index (*IFI*), network aggradation (*AGG*), network homogenization (*HMG*), and network amplification (*AMP*). Our results make two primary contributions. First, they demonstrate that five of the indicators—*FCI*, *Indirect/Direct*, *IFI*, *AGG* and *HMG*—are relatively robust to the flow and storage uncertainty in the Lake Lanier model. This stability lets us draw robust conclusions about the Lake Lanier ecosystem organization (e.g., phosphorus flux in the lake is dominated by internal processes) in spite of uncertainties in the model. Second, we show that the majority of the indicators co-vary and that most of their common variation could be mapped onto two latent factors, which we interpret as (1) system integration and (2) boundary influences.

Key words: aquatic ecosystem, environ analysis, flow analysis, indirect effects, network analysis, uncertainty

1 “The desire for simplification is justified, but the same does not always
2 apply to simplification itself, which is a working hypothesis, useful as long
3 as it is recognized as such and not mistaken for reality. The greater part of
4 historical and natural phenomena are [sic] not simple, or not simple in the
5 way that we would like.” (Levi, 1988)

6 **1 Introduction and Motivation**

7 Ecosystems are open, non-equilibrium, thermodynamic systems that are knit
8 together by an intricate network of energy, matter, and information exchanges
9 among biological organisms and their environments (Capra, 1996; Higashi and
10 Burns, 1991; Jørgensen, 2002; Ulanowicz, 1986). Like other types of complex
11 adaptive hierarchical systems, ecosystems appear to self-organize in response
12 to thermodynamic gradients (Levin, 1998; Müller, 1996; Schneider and Kay,
13 1994). However, a clear understanding of ecosystem organization and transfor-
14 mation has yet to crystalize, despite several empirical and theoretical attempts
15 (e.g., Gunderson and Holling, 2002; Jørgensen, 2002; Odum, 1969; Ulanowicz,
16 1986). This understanding is critical because ecosystems provide the natu-
17 ral capital and services that support human endeavors (Costanza et al., 1997;
18 Daily, 1997), and this knowledge is essential for effective ecosystem assessment
19 and management (Christensen et al., 1996; Reichman and Pulliam, 1996).

20 Ecosystem science has a long history of characterizing patterns of organization
21 and development (e.g., Lindeman, 1942; Margalef, 1963; Odum, 1969; Teal,

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22 1962). Numerous measures of ecosystem organization have been proposed in-
23 cluding gross production-to-biomass ratio, species diversity, flow diversity, cy-
24 cling (Odum, 1969), power (Lotka, 1922), and ascendancy (Ulanowicz, 1986,
25 1997). As systems develop, several of these measures appear to consistently
26 increase or decrease, suggesting possible orienting forces or constraints on self-
27 organization (Fath et al., 2001; Müller and Leupelt, 1998; Odum, 1969; Patten,
28 1998; Schneider and Kay, 1994; Ulanowicz, 1986).

29 In Jørgensen’s (2002) efforts to weave together multiple ecosystem theories
30 into a coherent whole, he acknowledged that a plurality of approaches re-
31 mains necessary. Holoecology (Patten, in prep.) offers one distinct approach.
32 The overarching goals of the Holoecology Research Program are (1) to un-
33 derstand the lawful processes that create, constrain, and sustain ecological
34 systems and (2) to create a formal theory of environment. Network Environ-
35 Analysis (NEA) is the primary methodology of Holoecology (Fath and Pat-
36 ten, 1999b; Matis and Patten, 1981; Patten, in prep.; Patten et al., 1976), and
37 it is fundamentally an environmental application and extension of economic
38 Input–Output Analysis (Leontief, 1966). Although it has elements in com-
39 mon with other forms of ecosystem network analysis such as Input–Output
40 Analysis as implemented in Ecopath (Christensen and Pauly, 1992; Chris-
41 tensen and Walters, 2004) or Acendency Theory (Allesina and Bondavalli,
42 2004; Ulanowicz, 1986, 1997), NEA is distinguished by its explicit environ-
43 ment focus. It is used in the holistic study of ecological networks to describe
44 and quantify component-level, bounded environments within systems, termed
45 environs (Patten, 1978, 1981, 1982).

46 A strength of NEA is its inclusion of several indicators that summarize whole-
47 system organization including total system storage (TSS), total boundary
48 input or output ($Boundary$), total system throughflow (TST), Finn cycling

49 index (*FCI*), average path length or network aggradation (*AGG*), ratio of
50 indirect-to-direct flows (*Indirect/Direct*), indirect flow index (*IFI*), network
51 homogenization (*HMG*), and network amplification (*AMP*). *TSS*, *Boundary*,
52 *TST*, *FCI*, and *AGG* are commonly used in other forms of ecosystem network
53 analysis, while *Indirect/Direct*, *IFI*, *HMG*, and *AMP* are unique to NEA
54 (Fath and Patten, 1999b; Patten, 1998). Collectively, these indicators provide
55 insight into the flow and storage organization of ecosystem models. Based on
56 these indicators, Patten (1998, in prep.) argues that ecological systems are
57 integrated by indirect effects.

58 The Achilles heel of these indicators, NEA, and ecosystem network analyses in
59 general is that they are based on models of real ecosystems. Ecological model-
60 ing remains a challenging endeavor with no consensus on the best approach or
61 evaluation procedures (e.g., Ginzburg and Jensen, 2003; Jakeman et al., 2006;
62 Oreskes et al., 1994). In truth, an infinite number of models fit a given data
63 set, and choosing among them remains difficult. As the opening quote from
64 Levi states, natural phenomena are not always simple in the way we wish.

65 Previous research found that ecosystem indicators can be highly dependent
66 upon the model form, especially the model aggregation (e.g., Abarca-Arenas
67 and Ulanowicz, 2002; Cale and Odell, 1979; Cale and O'Neill, 1988; Gardner
68 et al., 1982; Pahl-Wostl, 1992; Sugihara et al., 1997). Less research has explored
69 the sensitivity of these indicators to uncertainty in the magnitude of flows and
70 storages (but see Allesina and Bondavalli, 2003; Bosserman, 1983; Fath, 2004),
71 which can arise from both incomplete or unknown data for model calibration
72 and from evaluation or empirical measurement error.

73 In this work, we determined the sensitivity and interrelations of the indicators
74 of ecosystem organization listed above to flux and storage uncertainty in a

75 phosphorus-based ecosystem model of Lake Sidney Lanier, USA. Specifically,
76 we investigated two hypotheses. First, we conjectured that six of the focal sys-
77 tem indicators (*FCI*, *Indirect/Direct*, *IFI*, *AGG*, *AMP*, and *HMG*) would
78 be robust to model flux uncertainty. Quantitatively we expected each of these
79 indicators to be less variable than *TST* and *TSS*; qualitatively we expected
80 their interpretations to be consistent. Support for this hypothesis lets us draw
81 more robust conclusions about Lake Lanier despite uncertainty in the model
82 uncertainty. Second, we anticipated that these six indicators would charac-
83 terize different aspects of the same latent factor, which we tentatively term
84 *system integration*. This hypothesis implies that the indicators will be highly
85 correlated. We also expected that they would largely map into one principle
86 latent factor. Support for this hypothesis would let us reduce the number of
87 NEA indicators required to characterize the degree of system integration by
88 indirect effects.

89 **2 Materials and Methods**

90 *2.1 Study System*

91 Lake Sydney Lanier is a large reservoir in the headwaters of the Chattahoochee
92 River in Georgia, USA that drains a 2704 km² watershed, and at the conserva-
93 tion pool elevation of 326 m it covers 150 km² with 869 km of shoreline (Fath
94 and Beck, 2005; Guan, 1993). Constructed in the 1950s by the U.S. Army
95 Corp of Engineers, the reservoir was initially to provide flood control, hydro-
96 electric power, and downstream navigation regulation (U.S. Army Corps of
97 Engineers, 2005). However, the reservoir lies just northeast of downtown At-
98 lanta, one of the fastest growing metropolitan areas in the United States for

99 the past two decades (Metro Atlanta Chamber of Commerce, 2000), and it
100 has become an increasingly important source of drinking water, wastewater
101 dilution, and recreation for the region. Lake Lanier is a key resource support-
102 ing the expansion of Atlanta and the rapid urbanization of its own watershed
103 (Fath and Beck, 2005).

104 *2.2 Model Construction*

105 To address our hypotheses, we required a model of phosphorus flux in the Lake
106 Lanier ecosystem that was valid given our knowledge of the system, but that
107 let us investigate the space of plausible variability. We constructed this model
108 in two steps. We first built a conceptual model modified from an existing
109 model of the system. We then identified a set of plausible parameterizations
110 for this model whose generated behavior matched the empirically observed
111 summer behavior of the ecosystem.

For our model, we modified the thirteen compartment Lanier ecosystem model introduced by (Osidele and Beck, 2004). We converted the original model to a consistent currency (phosphorus) and replaced the nonlinear mechanistic functions with phenomenological donor controlled functions, which we modeled with first-order differential equations of the generic form

$$\frac{dX}{dt} = -\lambda_X \cdot X, \quad (1)$$

112 where X is a generic compartmental state variable, λ_X is a rate constant,
113 and t is continuous time. Since the function is donor controlled, the negative
114 sign indicates a loss or transfer from the donor compartment. We made these
115 model changes so that the output would readily meet the assumptions of NEA
116 described in Section 2.3.

117 The modified model shown in Figure 1 has eleven state variables or storage
118 compartments (mg P m^{-2}), twenty six within-system flows ($\text{mg P m}^{-2} \text{ d}^{-1}$),
119 five boundary inputs, and 4 boundary losses. The compartments represent (1)
120 epilimnion soluble reactive phosphorus (SRP), (2) hypolimnion SRP, (3) phy-
121 toplankton, (4) microbes (bacteria, etc.), (5) microzooplankton, (6) macrozoo-
122 plankton, (7) macroinvertebrates (insect larvae, etc.), (8) larval–juvenile fish,
123 (9) suspended detritus, (10) sediment detritus, and (11) sediment pore space
124 phosphorus. Phosphorus enters the model via fluxes into compartments 1, 2,
125 8, 9 and 11, and exits from compartments 1, 3, 7, and 8.

126 Given this system conceptualization, we then used Monte Carlo simulations
127 (Manly, 1991) and regionalized sensitivity analysis (Osidele and Beck, 2001;
128 Spear and Hornberger, 1980) to identify parameterizations whose summer
129 behavior of epilimnion phosphorus, phytoplankton, and fish fell within their
130 empirically known variability. We defined the behavior targets as a range of
131 values to account for uncertainty and spatial variability in the observational
132 data, which we obtained from the literature for phosphorus and phytoplankton
133 (Hatcher et al., 1994), and fish (Weaver, 2000). These records indicate that
134 the average annual phosphorus concentration in the photic zone ranges from 2
135 to $4 \mu\text{g L}^{-1}$, with no distinct spatial pattern of variation across the reservoir.
136 Phytoplankton chlorophyll concentrations range from $7 \mu\text{g L}^{-1}$ at the tributary
137 inlets to $2 \mu\text{g L}^{-1}$ at the dam site, and larval–juvenile fish average 20 to
138 50 kg ha^{-1} lake wide. We then translated these observations into the model
139 currency to prescribe the following summer behavior definition:

- 140 • epilimnion SRP concentration should be between 58 and 70 mg P m^{-2} ;
- 141 • phytoplankton concentration should be between 13 and 52 mg P m^{-2} ;
- 142 • larval–juvenile fish concentration should be between 100 and 250 mg P m^{-2} .

143 The regionalized sensitivity analysis considered 30 parameters (i.e., the mass
144 transfer rate constants λ_X in equation 1). The program sampled each param-
145 eter from a uniform distribution over the interval $[0.0, 0.5]$, i.e., 0% to 50%
146 rate of loss from each donor compartment. We considered parameterizations
147 that satisfied the summer behavior definition to be plausible representations
148 given our knowledge of the system. Furthermore, all successful parameteri-
149 zations generated static, steady-state behavior. To insure that the sample of
150 parameterizations represented the larger population, we compared the mean
151 and variance of the ecosystem indicators in successful samples from trial runs
152 of 500, 1000, and 5000 realizations. We used a Welsh t-test to compare the
153 means as it does not assume identical variance (Dalgaard, 2002).

154 *2.3 Network Environ Analysis—Throughflow*

155 NEA is a family of input–output methods that analytically decompose ob-
156 served flows and storages to identify their origins and fates within a system
157 of interest (Fath and Patten, 1999b; Patten et al., 1976). This methodology
158 includes analyses of structure, throughflow, storage, utility, and control within
159 systems, and is extensively described in the literature (e.g., Borrett and Pat-
160 ten, 2003; Fath and Borrett, 2006; Fath and Patten, 1999b; Gattie et al., 2006;
161 Matis and Patten, 1981; Patten et al., 1976). Hence, we only briefly summa-
162 rize the input and output variables of the output-oriented throughflow analysis
163 and the calculations for the indicators of interest.

164 NEA requires four input variables: (1) the observed flows from compartment
165 j to i ($\mathbf{F} = (f_{ij})$), (2) the boundary inputs (z_i), (3) the boundary outputs (y_i),
166 and (4) the storage values (x_i) of an n compartment system. This method-
167 ology assumes that all variables have a consistent currency (i.e., carbon or

168 phosphorus) and that the system they describe is in a static steady-state (i.e.,
 169 the inflows equal the outflows for each compartment). The static steady-state
 170 assumption is required for the current methods to partition the integral flows
 171 into categories based on pathway types (e.g., direct, indirect). As ecologists
 172 are often interested in ecosystem dynamics, this assumption limits the use of
 173 NEA (see discussion in Borrett et al., 2006; Fath and Patten, 1999b). How-
 174 ever, investigation of the static, steady-state case provides a snapshot of the
 175 system organization that is otherwise unavailable. This assumption influences
 176 the whole-system indicators because they depend on the throughflow decom-
 177 position, but we expect their qualitative interpretations to remain valid.

178 Given these variables, the output-oriented throughflow analysis characterizes
 179 the fate of material flowing into the system, which is accomplished with three
 180 analytical steps. First, we calculate the total material flowing through each
 181 node, which at steady-state is

$$\begin{aligned}
 \mathbf{T} = T_k^{(in)} &= \sum_{j(\neq k)1}^n f_{kj} + z_k \\
 &= T_k^{(out)} = \sum_{i(\neq k)1}^n f_{ik} + y_k.
 \end{aligned}
 \tag{2}$$

Second, we determine the dimensionless direct flow intensities from j to i as

$$\mathbf{G} = (g_{ij}) = f_{ij}/T_j.
 \tag{3}$$

182 Third, we find the dimensionless integral (boundary + direct + indirect) flow
 183 intensities

$$\begin{aligned}
\mathbf{N} &= \underbrace{\mathbf{I}}_{\text{Boundary}} + \underbrace{\mathbf{G}^1}_{\text{Direct}} + \underbrace{\mathbf{G}^2 + \dots + \mathbf{G}^m + \dots}_{\text{Indirect}} + \dots \\
&= (\mathbf{I} - \mathbf{G})^{-1},
\end{aligned}
\tag{4}$$

184 where $\mathbf{I} = \mathbf{G}^0$ is the matrix multiplicative identity and the elements of \mathbf{G}^m
185 are the flow intensities from j to i over all pathways of length m . We can then
186 verify that equation 2 is a true partition of the input flows across the many
187 pathways by ensuring that $\mathbf{T} = \mathbf{Nz}$. From these initial calculations, we then
188 derive the indicators of whole-system organization shown in Table 1.

189 2.4 Data Analysis

190 Our data analysis involved several steps. First, we calculated the whole-system
191 indicators for each plausible model parameterization using a modified version
192 of the MATLAB function NEA.m (Fath and Borrett, 2006). We then compared
193 the coefficient of variation for each indicator to evaluate robustness with the
194 assumption that more robust indicators are less variable. We chose to use the
195 dimensionless coefficient of variation because the indicator's absolute values
196 are measured on different scales. For example, TST has units of mg P m^{-2} ,
197 and $Indirect/Direct$ is a dimensionless ratio where $0 < Indirect/Direct < \infty$.
198 The variability of TST , TSS , and $Boundary$ provided a baseline indication
199 of the flow and storage variability in the alternative parameterizations.

200 We used two statistical approaches to determine whether the indicators char-
201 acterized different aspects of any underlying factors. We initially evaluated the
202 correlation strength of the indicators using ordinary least-squares regression.
203 Then, we detected latent or underlying variables with principle components

204 factor analysis (Grimm and Yarnold, 1995; Johnson, 1998), using the Kaiser
205 criterion to select the number of relevant factors. These statistical methods
206 assume that the variable distributions are normal and that the relationships
207 are linear. To meet these assumptions, we used the natural log transformation
208 of *TST*, *TSS*, *Indirect/Direct*, and *AGG*. We used a combination of MAT-
209 LAB (version 6.5, The Mathworks, Inc.) and R (R Development Core Team,
210 2005) for our calculations.

211 **3 Results**

212 There are three aspects to our results. We first show results of our model
213 identification and illustrate why the sample used for subsequent analysis is
214 representative of the space of valid flow–storage models. We then present the
215 ecosystem indicator variability within this sample of model parameterizations.
216 Finally, we describe relationships among the indicators.

217 *3.1 Model Identification*

218 With the methodology described in Section 2.2, we identified multiple model
219 parameterizations whose generated behavior satisfied our empirically derived
220 criteria for epilimnion phosphorus, phytoplankton, and fish storage (Figure 2).
221 Monte Carlo simulations with sample sizes of 500, 1000, and 5000 produced
222 14, 18, and 90 plausible parameterizations, respectively. To determine if these
223 samples represent the population of valid parameterizations, we compared the
224 mean and variability of the ecosystem indicators among the samples, shown in
225 Figure 3, and found no statistically significant differences between the samples
226 for *TST*, *TSS*, *FCI*, *Indirect/Direct*, *IFI*, *AGG*, and *AMP*. However, sam-

227 ple variance of the total boundary inputs (*Boundary*) is significantly different,
228 increasing with sample size, and the mean and variability of *HMG* is statis-
229 tically different between the 500 and 5000 trials. While a larger sample size
230 might have stabilized results for total boundary flow and homogenization, we
231 concluded that, for our purposes, the 90 plausible parameterizations provide a
232 large enough sample size to address our hypotheses. Therefore, all subsequent
233 analysis is of the 90 plausible parameterizations.

234 While each of the 90 parameterizations generates a valid model because they
235 satisfy the behavior criteria in Section 2.2, Table 2 shows that there remains
236 substantial uncertainty in the phosphorus flow and storage values. For exam-
237 ple, phosphorus in the sediment pore space had a mean of 341.3 (± 588.7).
238 Boundary flows were less variable with a maximum standard deviation of
239 21.5 associated with the largest mean boundary from the larval–juvenile fish
240 compartment. Internal system flows ranged from a minimum of 4.3 (± 2.7)
241 from phytoplankton to epilimnion P to a maximum of 55.8 (± 47.2) from the
242 hypolimnion to the microbial compartment. Notice that phosphorus stored in
243 the epilimnion, phytoplankton, and larval–juvenile fish varies the least because
244 these compartments are constrained by the empirical observations.

245 3.2 Indicator Variability

246 Table 3 reports the mean and variability of the NEA ecosystem indicators from
247 the 90 model parameterizations. The mean and standard deviation of *TST*,
248 *Boundary*, and *TSS* are 749 (± 303), 96 (± 20) and 1634 (± 985), respectively.
249 The mean *FCI* indicates that recycling contributed on average 39% of phos-
250 phorus *TST* in Lake Lanier. We expected a large degree of recycling because
251 we are analyzing an ecosystem model of phosphorus cycling. Furthermore, the

252 values of *IFI* and the *Indirect/Direct* suggest that a large fraction of *TST*
253 comes from indirect flows. The degrees of *AGG*, *HMG*, and *AMP* imply that
254 (1) an average input passes through an average of 7.84 nodes before exiting
255 the system, (2) the off-diagonal integral flow probabilities were 3 times more
256 evenly distributed than the direct flow probabilities, and (3) on average 22%
257 of the integral flow probabilities exceeded unity. This degree of amplification
258 indicates that generally two compartments received more than a unit input,
259 probably due to recycling.

260 To compare the relative variability of the ecosystem indicators, we calculated
261 their coefficients of variation (CV; Table 1). The CV of *TST* (0.40) and *TSS*
262 (0.60) reflect the whole system flow and storage variability due to model un-
263 certainty, so we used these values as benchmarks to compare the other seven
264 indicators. *IFI* and *HMG* have the lowest CVs at 0.10, while *AMP* varies
265 the most at 0.59. *AMP* is the only indicator to have a CV larger than *TST*,
266 close to *TSS*. The CV of *AGG* and *Indirect/Direct* are similar at 0.34 and
267 0.38 respectively, which is close to that of *TST*. *FCI* is less variable at 0.28,
268 and *Boundary* is 0.21. Given these quantitative results, the relative variability
269 and hence the robustness of the indicators is not uniform.

270 Despite their quantitative variability derived from model uncertainty, the qual-
271 itative interpretations of the indicators are robust. In all 90 parameterizations
272 we conclude that recycling was a large if not dominant source of phospho-
273 rus flux. *Indirect/Direct* always surpassed unity implying that indirect flows
274 were dominant, and network homogenization, amplification, and aggradation
275 always occurred.

277 Analysis of the statistical relationships among the ecosystem indicators re-
278 vealed several interesting patterns. We first describe the correlations among
279 the variables, and then explain the results of our factor analysis.

280 The pairwise scatter plots of the indicator combinations shown in Figure 4
281 reveal several close associations. In 27 of 36 cases there is a statistically sig-
282 nificant positive association between the variables (Table 4). For example,
283 strong positive associations exist between *FCI*, $\ln(\text{Indirect/Direct})$, *IFI*,
284 $\ln(\text{AGG})$, and *AMP*, whose correlation coefficients ranged from 0.89 to 0.98.
285 Both the scatter plots and correlation coefficients suggest that several of these
286 indicators may be influenced by the same underlying factor, lending support
287 to our second hypothesis. However, in some cases a linear model may not be
288 the most appropriate (e.g., between $\ln(\text{Indirect/Direct})$ and *IFI*). *Boundary*
289 and *HMG* provide exceptions to the trend. Although *Boundary* does increase
290 with $\ln(\text{TST})$ and $\ln(\text{TSS})$, it is not significantly correlated with the other
291 variables. *HMG* shows relatively weak correlations with the other indicators,
292 three of which are not statistically significant.

293 We used a principle component factor analysis to further evaluate these asso-
294 ciations. After examining several alternatives (not shown) we decided that two
295 factors are appropriate for the data, which is supported by the Kaiser crite-
296 rion. Only two correlation matrix eigenvalues are larger than unity ($\lambda_1 = 5.94$
297 and $\lambda_2 = 1.46$), which implies that only the first two factors account for more
298 variability than any single variable. Furthermore, the first factor captures 65%
299 of the variance and the cumulative variance accounted for by the two factors
300 was 80%. Including a third factor only increased this to 85%. The factor load-
301 ings shown in Table 5 indicate the correlation of each NEA indicator with

302 the two factors. Factor 1 is highly associated with the variability in $\ln(TST)$,
303 FCI , $\ln(Indirect/Direct)$, IFI , $\ln(AGG)$ and AMP . HMG , $\ln(TSS)$, and
304 $Boundary$ appear to have a small role in Factor 1. $Boundary$ dominates Fac-
305 tor 2, with minor contributions from $\ln(TST)$ and $\ln(TSS)$.

306 In contrast to principal components analysis, factor analysis only examines
307 the common variation of the variables (Tabachnick and Fidell, 1983). Indica-
308 tor uniqueness indicates the proportion of variation not in common with the
309 other variables (Table 5). HMG has the highest uniqueness factor, followed
310 by $\ln(TSS)$, and then AMP . The remaining variables had a uniqueness factor
311 below 0.10.

312 Figure 5 illustrates these results with a plot of the factor loadings. In this
313 plot, an indicator's distance from the origin indicates its strength of asso-
314 ciation with the factor represented by the axis. As $\ln(AGG)$ is the furthest
315 distance from the origin of the x-axis, it is the most highly associated with
316 Factor 1. This plot emphasizes the clustering of the variability of $\ln(AGG)$,
317 IFI , $\ln(Indirect/Direct)$, FCI , and AMP , and their distance from variation
318 in total boundary flux ($Boundary$). Three factors do not fall along an axis—
319 HMG , $\ln(TSS)$, and $\ln(TST)$ —which could make interpretation of the fac-
320 tors difficult. However, the uniqueness vector indicates that variation in HMG
321 and $\ln(TSS)$ is largely independent of the other indicators, and $\ln(TST)$ is a
322 special case that we address in the discussion.

323 4 Discussion

324 The results reported here generally support our hypotheses regarding indi-
325 cator robustness and their interrelationships, but they also suggest a more

326 complicated story than we initially expected. In this Section, we discuss the
327 evidence for each hypothesis, offer additional explanations for these results,
328 and interpret their significance for the growth and development of ecological
329 systems in general and Lake Lanier in particular.

330 4.1 *Indicator Robustness*

331 The NEA indicators of ecosystem organization are qualitatively consistent
332 and quantitatively differentially robust, lending support to our first hypoth-
333 esis. Qualitatively the index interpretations did not change. Indirect flows
334 consistently dominate direct, both indirect and cycled flows consistently com-
335 prise a large proportion of TST , and some degree of network aggradation,
336 network homogenization and network amplification occur. Quantitatively, the
337 indicators have differing degrees of robustness, decreasing in variability as
338 $AMP > Indirect/Direct > AGG > FCI > HMG = IFI$. Given the Lake
339 Lanier model structure and the relatively large flow and storage uncertainty,
340 the magnitudes of network homogenization and indirect flow index are re-
341 markably consistent.

342 We claim that the small variability in the ecosystem indicators lets us cir-
343 cumvent part of the modeling and data uncertainty to draw more robust con-
344 clusions regarding the condition of the Lake Lanier ecosystem. Although we
345 lack certainty about the true summer flux and storage of phosphorus for some
346 of the model compartments, we expect the values to occur within our model
347 parameterizations. Thus, the more constrained ecosystem indicators should
348 be informative of the real system organization. An alternative interpretation
349 is that an indicator's consistency implies that it is uninformative because it
350 lacks the necessary sensitivity. We doubt this is the case as previous appli-

351 cations of these indices demonstrate a wide range of values for these indices
352 (e.g., Borrett et al., 2006; Fath, 2004; Finn, 1980).

353 From this study, we draw two primary conclusions about the Lake Lanier
354 ecosystem organization. First, internal processes heavily influence phospho-
355 rus flow and storage. This is evident in the consistently high indirect flow
356 index, indirect-to-direct ratio, and degree of recycling. This result suggests a
357 relatively well developed ecosystem as the system gains high utility from the
358 phosphorus inputs. Second, based on the robust measure of network homoge-
359 nization we conclude that even though phosphorus storage is heterogeneous,
360 phosphorus in the flows is well mixed.

361 4.2 *Indicator Interrelations*

362 Our second hypothesis anticipated that the NEA indicators were different
363 measures of the same underlying factor, which we expected to be a form
364 of functional connectivity we termed *system integration*. The results revealed
365 many associations among the indicators and that their common variance could
366 be largely partitioned into two latent factors. We can chiefly attribute common
367 variance in six of the nine measures (*TST*, *FCI*, *Indirect/Direct*, *IFI*, *AGG*,
368 and *AMP*) to one latent factor, which suggests that they capture different
369 aspects of a common unmeasured system variable. In addition, we ascribe
370 total boundary flows (*Boundary*) and a portion of *TST* is attributed to a
371 second factor. Together these latent factors account for about 80% of the total
372 variance. These results support our hypothesis with one major exception: the
373 variation of *HMG* has a high uniqueness factor.

374 Further consideration of these indicators explains why the relationships exist
375 among the variables in Factor 1. From a given network topology there are two

376 ways of increasing TST . The first is to increase energy–matter input into the
377 system. This action increases TST , but in this case the organization of the
378 system remains constant. The second approach is to increase the system recy-
379 cling by reappportioning the flux distribution, which necessarily increases IFI
380 because by definition all cyclic flows are indirect¹. In addition, this recycling
381 likely increases *Indirect/Direct* (see Higashi and Patten, 1989; Patten, 1991,
382 for additional detail). Another consequence is that the proportion of TST de-
383 rived from *Boundary* will decline and its inverse, AGG , will increase. Likewise
384 AMP should increase because the proportion of compartments receiving more
385 flow than they contribute depends directly dependent upon recycling (Patten
386 et al., 1990). Thus, cycling plays a central role in each of these six indicators.

387 Our discovery that the nine ecosystem indicators cluster around two latent
388 factors denotes that they contain some redundancy, but we wonder if the
389 factors might have additional import. Grimm and Yarnold (1995) advise that
390 caution is wise when interpreting factor analysis, but our findings suggest that
391 we can link the two emergent factors to the conceptual model of ecosystem
392 growth introduced by Jørgensen et al. (2000) and extended by Fath et al.
393 (2004). These authors describe four forms of growth:

394 **Form 0—Boundary growth.** A constant source of low-entropy material
395 sustains open thermodynamic systems and fundamentally limits the work a
396 system can perform. This growth Form 0 represents the increase in bound-
397 ary input.

398 **Form I—Structural growth.** This growth type corresponds to increasing
399 biomass caused by component growth in number, size, and types.

400 **Form II—Network growth.** This form of growth captures shifts in the in-

¹ The converse is not necessarily true: all indirect flows are are not cyclic. This implies that $IFI \geq FCI$ and that the values will converge as cycling increases.

401 ternal organization of the system, which includes an increase in connectivity
402 and recycling of energy and matter.

403 **Form III—Informational growth** This type of growth augments the inter-
404 nal order of the informational content of the system, and includes behavioral
405 changes from a more exploitative to a more conservative system as well as
406 possible increases in genetic information of component organisms.

407 As cycling is the essential element linking *TST*, *FCI*, *Indirect/Direct*, *IFI*,
408 *AGG*, and *AMP* in Factor 1, we suspect that this Factor represents Growth
409 Form II. We also expect *HMG* to be associated with this growth type even
410 though its variability is largely independent. Furthermore, energy–matter cy-
411 cles are fundamental to system growth and development because they estab-
412 lish routes of cybernetic feedback (DeAngelis et al., 1986; Patten and Odum,
413 1981), and provide a means of integrating ecosystem flows and storage (Pat-
414 ten et al., 1990; Ulanowicz, 1983). Therefore, we tentatively suggest that this
415 factor is Patten’s hypothesized system integration through indirect effects.

416 Factor 2 captures both the variability of boundary flows and a portion of
417 the variability of *TST* and expresses the effects of environmental variability.
418 Therefore, this factor relates to Growth Form 0. Notice that *TST* is a com-
419 ponent of this factor as well as the first because boundary flows and internal
420 flows are combined in its calculation.

421 Variation in *TSS* has a large uniqueness factor, so it was only weakly associ-
422 ated with Factor 1 and Factor 2. By definition, *TSS* is an indicator of Growth
423 Form I.

424 5 Related Work

425 The work reported here builds upon previous developments in ecological net-
426 work analysis. In this section, we briefly describe related work that established
427 or suggested indicator interrelationships as well as one study that anticipated
428 the indicator robustness. We conclude by highlighting the unique contribution
429 of our work.

430 Several studies laid the foundations for the indicator interrelationship hypoth-
431 esis. Higashi (Higashi and Patten, 1986; Patten, 1991; Patten et al., 1990)
432 showed algebraically that increasing the number of nodes, connectance, stor-
433 age, strength of direct flows, or the amount of recycling tends to increase
434 *Indirect/Direct*. Hence, the positive association of this ratio with *TSS*, *TST*
435 and *FCI* is not surprising. Further, Christensen (1995) compared several
436 proposed network indicators of ecosystem maturity in a study of 41 aquatic
437 ecosystem models, which had a consistent currency but varied in the number of
438 nodes and connectance. He found a strong correlation between *FCI* and *AGG*,
439 but they were not well correlated with *TST*. In his principle components anal-
440 ysis, *FCI* and *AGG* were closely associated with his first component, while
441 *TST* was more closely associated with the second. This finding agrees with
442 our results, but ours shows a stronger relationship between *TST* and *FCI*
443 and *AGG*. The different model currencies likely drive this incongruity. With
444 a more analytical approach, Fath et al. (2001) used a five mode pathway de-
445 composition to show how simultaneous increases in *TST*, *TSS*, *Boundary* and
446 *FCI* were reconcilable. In their discussion of *AMP* and *HMG*, Patten et al.
447 (1990) illustrated how recycling influence both of these indicators. When Fath
448 and Patten (1999a) introduced the quantitative measure for *HMG*, they con-
449 firmed that it tended to increase with cycling in a twenty node model. Later,

450 Fath (2004) used large ($n > 100$) cyber-ecosystem models to show that both
451 *Indirect/Direct* and *HMG* tended to increase with *FCI*. However, network
452 amplification never occurred in these models, which may be attributed to the
453 restricted cycling range in the cyber-ecosystem models ($0.14 < FCI < 0.26$).
454 Borrett et al. (2006) observed a strong association between *Indirect/Direct*
455 and *AGG* in sixteen nitrogen network models of the Neuse River Estuary.
456 These networks were highly retentive and exhibited a large degree of cycling.
457 The first characteristic caused *Direct* to be nearly equivalent to *Boundary*
458 and the second characteristic made *Indirect* and *TST* to converge such that
459 $TST/Boundary \sim Indirect/Direct$.

460 Allesina and Bondavalli's (2004) study of the effects of alternative flow bal-
461 ancing procedures is the most closely related work to our investigation of the
462 indicator's robustness to uncertainty. They found that altering model coeffi-
463 cients $\pm 10\%$ led to small changes in the balanced system's TST ², such that
464 50% of the resultant TST values were less than 2.5% of their original value.
465 Thus, TST appears to be robust to these flow perturbations. Significantly, the
466 authors compared the TST of a flow model that is slightly out of steady-state
467 to a balanced one. In contrast, all 90 model parameterizations used in our
468 study generated steady-state behavior. Furthermore, our investigation covers
469 a wider range of network indicators whereas Allesina and Bondavalli (2003)
470 restrict their investigation to TST and Ascendency, which is not in our study.

471 We conclude that several of the relationships among the NEA indicators were
472 known or anticipated from independent investigations. Our work brings these
473 indicators together into one quantitative study and begins to describe the
474 quantitative forms of these relationships. In addition, our data support the

² These authors calculate TST by summing both inputs and outputs in their calculation, whereas we include only one direction of these boundary flows because total inputs equal total outputs in a steady-state system.

475 hypotheses that many of these indicators are characterizing similar aspects of
476 ecosystem organization and are robust under flow and storage uncertainty.

477 **6 Future Work**

478 This research is a step toward understanding the effect of system uncertainty
479 on the NEA ecosystem indicators, but additional work is necessary to eval-
480 uate the generality of the results and to determine their significance for en-
481 vironmental impact assessment and management. We must be careful not to
482 overgeneralize from this case study. Our approach reveals the influence of flow
483 and storage uncertainty on the NEA indicators in Lake Lanier given the ini-
484 tial conceptual model. An important next step is to investigate the effect of
485 uncertainty in the model structure (e.g., conceptual model, flow formulation).
486 Further, we require additional case studies for two reasons. First, our anal-
487 ysis of indicator relationships is limited by their small range, which results
488 from their robustness. Second, we ultimately want to use these indicators to
489 characterize the Lake Lanier ecosystem’s healthy, maturity, or integrity. These
490 classifications are relative and require meaningful comparisons that do not yet
491 exist. More case studies will let us develop a meaningful scale to evaluate
492 ecosystem condition in the future.

493 **7 Concluding Remarks**

494 The results of this study make two primary contributions to ecosystem network
495 analysis. First, they demonstrate that five of the ecosystem network analysis
496 indicators—*FCI*, *Indirect/Direct*, *IFI*, *AGG* and *HMG*—are robust to the
497 flow and storage uncertainty in the Lake Lanier phosphorus model. This find-

498 ing lets us bypass the modeling problem and conclude that internal processes
499 heavily influence phosphorus flux in the Lake Lanier ecosystem. Second, the
500 results show that the majority of the nine ecosystem indicators investigated
501 co-varied and that their common variation could largely be mapped into two
502 latent factors, which we have tentatively interpreted as (1) system integration
503 (Growth Form II) and (2) boundary (Growth Form 0). This case study pro-
504 vides a window into the workings of network environ analysis and the Lake
505 Lanier ecosystem. Continued research into ecological network analysis sensi-
506 tivity to model uncertainty will lead to a better understanding of when our
507 model simplifications are justified, and when we must alter our simplifications
508 to better match natural phenomena.

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515 **References**

- 516 Abarca-Arenas, L. G., Ulanowicz, R. E., 2002. The effects of taxonomic ag-
517 gregation on network analysis. *Ecol. Model.* 149, 285–296.
- 518 Allesina, S., Bondavalli, C., 2003. Steady state of ecosystem flow networks: A
519 comparison between balancing procedures. *Ecol. Model.* 165, 221–229.
- 520 Allesina, S., Bondavalli, C., 2004. Wand: An ecological network analysis user-
521 friendly tool. *Environmental Modelling & Software* 19, 337–340.
- 522 Borrett, S. R., Patten, B. C., 2003. Structure of pathways in ecological net-

- 523 works: Relationships between length and number. *Ecol. Model.* 170, 173–
524 184.
- 525 Borrett, S. R., Whipple, S. J., Patten, B. C., Christian, R. R., 2006. Indi-
526 rect effects and distributed control in ecosystems. Temporal variability of
527 indirect effects in a seven-compartment model of nitrogen flow in the Neuse
528 River Estuary (USA)—Time series analysis. *Ecol. Model.* 194, 178–188.
- 529 Bosserman, R. W., 1983. Flow analysis sensitivities for models of energy or
530 material flow. *Bull. Math. Bio.* 45, 807–826.
- 531 Cale, W. G., Odell, P. L., 1979. Concerning aggregation in ecosystem models.
532 In: Halfon, E. (Ed.), *Theoretical Systems Ecology*. Academic Press, New
533 York, pp. 55–77.
- 534 Cale, W. G., O’Neill, R. V., 1988. Aggregation and consistency problems in
535 theoretical models of exploitative resource competition. *Ecol. Model.* 40,
536 97–107.
- 537 Capra, F., 1996. *The Web of Life: A New Scientific Understanding of Living*
538 *Systems*. Anchor Books, New York.
- 539 Christensen, N. L., Bartuska, A. M., Brown, J. H., Carpenter, S., D’Antonio,
540 C., Francis, R., Franklin, J. F., MacMahon, J. A., Noss, R. F., Parsons,
541 D. J., Peterson, C. H., Turner, M. G., Woodmansee, R. G., 1996. The report
542 of the Ecological Society of America committee on the scientific basis for
543 ecosystem management. *Ecological Applications* 6, 665–691.
- 544 Christensen, V., 1995. Ecosystem maturity—towards quantification. *Ecol.*
545 *Model.* 77, 3–32.
- 546 Christensen, V., Pauly, D., 1992. Ecopath-II—a software for balancing steady-
547 state ecosystem models and calculating network characteristics. *Ecol.*
548 *Model.* 61, 169–185.
- 549 Christensen, V., Walters, C. J., 2004. Ecopath with Ecosim: Methods, capa-
550 bilities and limitations. *Ecol. Model.* 172, 109–139.
- 551 Costanza, R., d’Arge, R., de Groot, R., Farber, S., Grasso, M., Hannon, B.,
552 Limburg, K., Naeem, S., O’Neill, R. V., Paruelo, J., Raskin, R. G., Sutton,
553 P., van den Belt, M., 1997. The value of the world’s ecosystem services and
554 natural capital. *Nature* 387, 253–260.
- 555 Daily, G. C. (Ed.), 1997. *Nature’s Services*. Island Press, Washington, D.C.
- 556 Dalgaard, P., 2002. *Introductory Statistics with R*. Springer, New York.
- 557 DeAngelis, D. L., Post, W. M., Travis, C. C., 1986. *Positive Feedback in*
558 *Natural Systems*. Springer-Verlag, New York.
- 559 Fath, B. D., 2004. Network analysis applied to large-scale cyber-ecosystems.
560 *Ecol. Model.* 171, 329–337.
- 561 Fath, B. D., Beck, M. B., 2005. Elucidating public perceptions of environ-
562 mental behavior: A case study of Lake Lanier. *Environmental Modelling &*
563 *Software* 20, 485–498.
- 564 Fath, B. D., Borrett, S. R., 2006. A MATLAB© function for network environ-
565 mental analysis. *Environmental Modelling & Software* 21, 375–405.
- 566 Fath, B. D., Jørgensen, S. E., Patten, B. C., Straškraba, M., 2004. Ecosystem

567 growth and development. *Biosystems* 77, 213–228.

568 Fath, B. D., Patten, B. C., 1999a. Quantifying resource homogenization using
569 network flow analysis. *Ecol. Model.* 107, 193–205.

570 Fath, B. D., Patten, B. C., 1999b. Review of the foundations of network environ-
571 iron analysis. *Ecosystems* 2, 167–179.

572 Fath, B. D., Patten, B. C., Choi, J. S., 2001. Complementarity of ecological
573 goal functions. *J. Theor. Biol.* 208, 493–506.

574 Finn, J. T., 1976. Measures of ecosystem structure and function derived from
575 analysis of flows. *J. Theor. Biol.* 56, 363–380.

576 Finn, J. T., 1980. Flow analysis of models of the Hubbard Brook ecosystem.
577 *Ecology* 61, 562–571.

578 Gardner, R. H., Cale, W. G., O’Neill, R. V., 1982. Robust analysis of aggrega-
579 tion error. *Ecology* 63, 1771–1779.

580 Gattie, D. K., Schramski, J. R., Borrett, S. R., Patten, B. C., Bata, S. A.,
581 Whipple, S. J., 2006. Indirect effects and distributed control in ecosystems.
582 Network environ analysis of a seven-compartment model of nitrogen flow in
583 the Neuse River Estuary (USA)—Steady-state analysis. *Ecol. Model.* 194,
584 162–177.

585 Ginzburg, L. R., Jensen, C. X. J., 2003. Rules of thumb for judging ecological
586 theories. *Trends in Ecology & Evolution* 19, 121–126.

587 Grimm, L. G., Yarnold, P. R., 1995. *Reading and Understanding Multivariate*
588 *Statistics*. American Psychological Association, Washington, D.C.

589 Guan, W., 1993. *Integrating Water Quality Modeling with Geographic Informa-*
590 *tion System—Application to Lake Sidney Lanier*. Doctoral dissertation,
591 University of Georgia.

592 Gunderson, L. H., Holling, C. S. (Eds.), 2002. *Panarchy: Understanding Trans-*
593 *formations in Human and Natural Systems*. Island Press, Washington, DC.

594 Han, B. P., 1997. On several measures concerning flow variables in ecosystems.
595 *Ecol. Model.* 104, 289–302.

596 Hatcher, K. J., Callahan, M. A., Nearing, M. A., Pancorbo, O., Patten, B. C.,
597 Rogers, L. F., Sellers, J., Van Den Avyle, M. J., 1994. *Diagnostic/Feasibility*
598 *Study of Lake Sidney Lanier, Georgia*. Project completion report, Prepared
599 for Georgia Environmental Protection Division.

600 Higashi, M., Burns, T. P. (Eds.), 1991. *Theoretical Studies of Ecosystems:*
601 *The Network Perspective*. Cambridge University Press, Cambridge.

602 Higashi, M., Patten, B. C., 1986. Further aspects of the analysis of indirect
603 effects in ecosystems. *Ecol. Model.* 31, 69–77.

604 Higashi, M., Patten, B. C., 1989. Dominance of indirect causality in ecosys-
605 tems. *Am. Nat.* 133, 288–302.

606 Jakeman, A. J., Letcher, R. A., Norton, J. P., 2006. Ten iterative steps in
607 development and evaluation of environmental models. *Environmental Mod-*
608 *elling & Software* 21, 602–614.

609 Johnson, D. E., 1998. *Applied Multivariate Methods for Data Analysts*.
610 Duxbury Press, Pacific Grove, CA.

611 Jørgensen, S. E., 2002. *Integration of Ecosystem Theories: A Pattern*. Kluwer

- 612 Academic Publishers, Boston.
- 613 Jørgensen, S. E., Patten, B. C., Straškraba, M., 2000. Ecosystems emerging:
614 4. Growth. *Ecol. Model.* 126, 249–284.
- 615 Leontief, W. W., 1966. *Input–Output Economics*. Oxford University Press,
616 New York.
- 617 Levi, P., 1988. *The Drowned and the Saved*. Summit Books, New York.
- 618 Levin, S. A., 1998. Ecosystems and the biosphere as complex adaptive systems.
619 *Ecosystems* 1, 431–436.
- 620 Lindeman, R. L., 1942. The trophic-dynamic aspect of ecology. *Ecology* 23,
621 399–418.
- 622 Lotka, A. J., 1922. Contribution to the energetics of evolution. *Proc. Natl.*
623 *Acad. Sci. USA* 8, 147–151.
- 624 Manly, B. F. J., 1991. *Randomization and Monte Carlo Methods in Biology*.
625 Chapman and Hall, New York.
- 626 Margalef, R., 1963. Certain unifying principles in ecology. *Am. Nat.* 97, 357–
627 374.
- 628 Matis, J. H., Patten, B. C., 1981. Environ analysis of linear compartmental
629 systems: The static, time invariant case. *Bull. Int. Stat. Inst.* 48, 527–565.
- 630 Metro Atlanta Chamber of Commerce, 2000. Clean water initiative final re-
631 port. Technical report, Metro Atlanta Chapter of Commerce.
- 632 Müller, F., 1996. Emergent properties of ecosystems—Consequences of self-
633 organizing processes? *Senckenbergiana maritima* 27, 151–168.
- 634 Müller, F., Leupelt, M. (Eds.), 1998. *Eco Targets, Goal Functions, and Ori-*
635 *entors*. Springer, New York.
- 636 Odum, E. P., 1969. The strategy of ecosystem development. *Science* 164, 262–
637 270.
- 638 Odum, H. T., 1962. Quantitative classification and limits of ecosystems. *Am.*
639 *Zoo.* 2, 434–434.
- 640 Oreskes, N., Shraderfrechette, K., Belitz, K., 1994. Verification, validation,
641 and confirmation of numerical-models in the earth-sciences. *Science* 263,
642 641–646.
- 643 Osidele, O. O., Beck, M. B., 2001. Identification of model structure for aquatic
644 ecosystems using regionalized sensitivity analysis. *Water Science and Tech-*
645 *nology* 43, 271–278.
- 646 Osidele, O. O., Beck, M. B., 2004. Food web modelling for investigating ecosys-
647 tem behaviour in large reservoirs of the South-eastern United States: Lessons
648 from Lake Lanier, Georgia. *Ecol. Model.* 173, 129–158.
- 649 Pahl-Wostl, C., 1992. The possible effects of aggregation on the quantitative
650 interpretation of flow patterns in ecological networks. *Mathematical Bio-*
651 *sciences* 112, 177–183.
- 652 Patten, B. C., 1978. Systems approach to the concept of environment. *Ohio*
653 *J. Sci.* 78, 206–222.
- 654 Patten, B. C., 1981. Environs: The superniches of ecosystems. *American Zo-*
655 *ologist* 21, 845–852.
- 656 Patten, B. C., 1982. Environs: Relativistic elementary particles for ecology.

657 Am. Nat. 119, 179–219.

658 Patten, B. C., 1991. Network ecology: Indirect determination of the life–
659 environment relationship in ecosystems. In: Higashi, M., Burns, T. (Eds.),
660 Theoretical Studies of Ecosystems: The Network Perspective. Cambridge
661 University Press, New York, pp. 288–351.

662 Patten, B. C., 1998. Network orientors: Steps toward a cosmography of ecosys-
663 tems: Orientors for directional development, self-organization, and autoevo-
664 lution. In: Müller, F., Leupelt, M. (Eds.), Eco Targets, Goal Functions, and
665 Orientors. Springer, Berlin, pp. 137–160.

666 Patten, B. C., in prep. Holoecology: The Unification of Nature by Network
667 Indirect Effects. Columbia University Press, New York.

668 Patten, B. C., Bosserman, R. W., Finn, J. T., Cale, W. G., 1976. Propaga-
669 tion of cause in ecosystems. In: Patten, B. C. (Ed.), Systems Analysis and
670 Simulation in Ecology, Vol. IV. Academic Press, New York, pp. 457–579.

671 Patten, B. C., Higashi, M., Burns, T. P., 1990. Trophic dynamics in ecosystem
672 networks: Significance of cycles and storage. *Ecol. Model.* 51, 1–28.

673 Patten, B. C., Odum, E. P., 1981. The cybernetic nature of ecosystems. *Ameri-
674 can Naturalist* 118, 886–895.

675 R Development Core Team, 2005. R: A language and environment for statisti-
676 cal computing. R Foundation for Statistical Computing, Vienna, Austria.
677 URL <http://www.R-project.org>

678 Reichman, O. J., Pulliam, H. R., 1996. The scientific basis for ecosystem man-
679 agement. *Ecological Applications* 6, 694–696.

680 Samuelson, P. A., 1948. *Economics: An Introductory Analysis*. McGraw–Hill
681 Book Co., New York, New York.

682 Schneider, E. D., Kay, J. J., 1994. Life as a manifestation of the second law
683 of thermodynamics. *Mathematical and Computer Modelling* 19, 25–48.

684 Spear, R. C., Hornberger, G. M., 1980. Eutrophication in Peel Inlet—2. Iden-
685 tification of critical uncertainties via generalized sensitivity analysis. *Wat.
686 Res.* 14, 43–49.

687 Sugihara, G., Bersier, L. F., Schoenly, K., 1997. Effects of taxonomic and
688 trophic aggregation on food web properties. *Oecologia* 112, 272–284.

689 Tabachnick, B. G., Fidell, L. S., 1983. *Using Multivariate Statistics*. Harper
690 & Row, New York.

691 Teal, J. M., 1962. Energy-flow in salt-marsh ecosystem of Georgia. *Ecology*
692 43, 614–624.

693 Ulanowicz, R. E., 1983. Identifying the structure of cycling in ecosystems.
694 *Mathematical Biosciences* 65, 219–237.

695 Ulanowicz, R. E., 1986. *Growth and Development: Ecosystems Phenomenol-
696 ogy*. Springer–Verlag, New York.

697 Ulanowicz, R. E., 1997. *Ecology, the Ascendent Perspective*. Columbia Uni-
698 versity Press, New York.

699 U.S. Army Corps of Engineers, 2005. Lake Sidney Lanier, February 21, 2005.
700 URL <http://lanier.sam.usace.army.mil/>

701 Weaver, O. R., 2000. Lake Lanier annual report, 1999. Technical report, Geor-

702 gia Department of Natural Resources, Fisheries Section of the Wildlife Re-
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Table 1. Network Environ Analysis indicators of whole-system organization.

Indicator	Symbol	Description	Formula
Total System Throughflow	TST	Sum of total flow into or out of nodes	$\sum_{k=1}^n T_k = \sum_{k=1}^n \sum_{k=1}^n (f_{kj} + z_k) = \sum_{k=1}^n \sum_{k=1}^n (f_{ik} + y_k)$
Total System Storage	TSS	Total amount of model currency stored in nodes	$\sum_{k=1}^n x_k$
Total Boundary Flow	$Boundary$	Total amount of boundary input or output	$\sum_{i=1}^n z_i = \sum_{j=1}^n y_j$
Finn Cycling Index	FCI	Cyclic portion of TST	$\sum_{i=1}^n ((n_{ii} - 1)z_i)$
Indirect/Direct	$Indirect/Direct$	Ratio of indirect to direct flow	$\frac{\sum (\mathbf{N} - \mathbf{I} - \mathbf{G})\mathbf{z}}{\sum \mathbf{G}\mathbf{z}}$
Indirect Flow Index	IFI	Proportion of TST derived from indirect flows	$\frac{\sum (\mathbf{N} - \mathbf{I} - \mathbf{G})\mathbf{z}}{TST}$
Homogenization	HMG	Tendency to uniformly distribute causality across the network	$\frac{CV(\mathbf{G})}{CV(\mathbf{N})}$
Amplification	AMP	Proportion of flows obtaining more than face value	$\frac{\#n_{ij} > 1 \quad (i \neq j)}{n(n-1)}$
Aggradation ^a	AGG	Average number of times an average input passes through the system	$\frac{TST}{Boundary}$

^a Aggradation is also known as average path length (Finn, 1976), flow multiplying ability (Han, 1997), and multiplier effect (Samuelson, 1948).

Table 2. Average (\pm SD) inter-compartment flows (\mathbf{F}) oriented from column to row, inputs (\mathbf{z}), outputs (\mathbf{y}) in mg P m⁻² d⁻¹, and storage (\mathbf{x} , mg P m⁻²) for 90 plausible models of phosphorus flow and storage in Lake Lanier. Compartment labels are in the vector Names.

$\mathbf{F} =$	0	33.6 (10.1)	4.3 (2.7)	0	0	0	0	0	0	0	0	0
	0	0	0	28.3 (29.6)	0	0	0	0	0	0	0	39.9 (25.8)
	25.3 (5.3)	0	0	0	0	0	0	0	0	0	0	0
	0	55.8 (47.2)	0	0	0	0	0	0	0	33.9 (28.4)	0	0
	0	0	5.2 (3.1)	29.6 (25.2)	0	0	0	0	0	38.2 (32.4)	0	0
	0	0	5.1 (3.3)	0	16.8 (15.1)	0	0	0	0	35.7 (24.0)	0	0
	0	0	0	0	18.8 (16.7)	0	0	0	0	38.3 (25.4)	0	0
	0	0	0	0	18.1 (13.9)	28.4 (17.2)	18.8 (16.0)	0	0	0	0	0
	0	0	5.4 (3.2)	31.7 (29.1)	19.3 (18.1)	29.1 (25.6)	19.2 (16.9)	39.8 (25.7)	0	0	0	0
	0	0	0	0	0	0	0	0	0	26.2 (23.3)	0	0
	0	0	0	0	0	0	0	0	0	0	26.2 (23.3)	0

Names =	Epilimnion P	$\mathbf{z} =$	6.9 (5.3)	$\mathbf{y}^T =$	19.4 (9.1)	$\mathbf{x} =$	64.4 (3.5)
	Hypolimnion P		21.1 (9.9)		0.0		228.5 (208.7)
	Phytoplankton		0		5.3 (2.9)		22.3 (7.0)
	Microbes		0		0.0		129.9 (106.7)
	Microzooplankton		0		0.0		77.6 (50.1)
	Macrozooplankton		0		0.0		154.0 (223.0)
	Macroinverts		0		19.1 (16.0)		81.4 (55.8)
	Larval/Juvenile fish		27.5 (12.6)		53.0 (21.5)		171.1 (42.5)
	Suspended detritus		27.6 (11.2)		0.0		152.6 (83.5)
	Sediment detritus		0		0.0		240.0 (398.4)
	Pore Space P		13.8 (8.6)		0.0		341.3 (588.7)

Table 3

Mean, standard deviation (SD), and coefficient of variation (CV) of Network Environment Analysis ecosystem indicators to flow and storage uncertainty in a phosphorus model of Lake Lanier. They are ordered by decreasing variability described by CV

Indicator	Mean	SD	CV
<i>TSS</i>	1634	985	0.60
<i>AMP</i>	0.21	0.12	0.59
<i>TST</i>	749	303	0.40
<i>Indirect/Direct</i>	7.45	2.83	0.38
<i>AGG</i>	7.84	2.69	0.34
<i>FCI</i>	0.39	0.11	0.28
<i>Boundary</i>	96	20	0.21
<i>HMG</i>	3.10	0.31	0.10
<i>IFI</i>	0.75	0.07	0.10

Table 4
 Pairwise linear ordinary least-squares regression coefficients ($Y = b + mX$).

<i>Y</i>	<i>X</i>	<i>b</i>	<i>m</i>	<i>Pr(> F)</i>
ln(<i>TST</i>)	ln(<i>TSS</i>)	2.94 *	0.50 *	0.00 *
ln(<i>TST</i>)	<i>Boundary</i>	5.78 *	0.01 *	0.00 *
ln(<i>TST</i>)	<i>FCI</i>	5.53 *	2.63 *	0.00 *
ln(<i>TST</i>)	ln(<i>Indirect/Direct</i>)	5.01 *	0.80 *	0.00 *
ln(<i>TST</i>)	<i>IFI</i>	3.62 *	3.94 *	0.00 *
ln(<i>TST</i>)	ln(<i>AGG</i>)	4.75 *	0.90 *	0.00 *
ln(<i>TST</i>)	<i>HMG</i>	5.88 *	0.23	0.07
ln(<i>TST</i>)	<i>AMP</i>	6.09 *	2.22 *	0.00 *
ln(<i>TSS</i>)	<i>Boundary</i>	6.75 *	0.01 *	0.03 *
ln(<i>TSS</i>)	<i>FCI</i>	6.41 *	2.20 *	0.00 *
ln(<i>TSS</i>)	ln(<i>Indirect/Direct</i>)	5.87 *	0.72 *	0.00 *
ln(<i>TSS</i>)	<i>IFI</i>	4.44 *	3.78 *	0.00 *
ln(<i>TSS</i>)	ln(<i>AGG</i>)	5.48 *	0.89 *	0.00 *
ln(<i>TSS</i>)	<i>HMG</i>	5.64 *	0.54 *	0.00 *
ln(<i>TSS</i>)	<i>AMP</i>	6.92 *	1.66 *	0.00 *
<i>Boundary</i>	<i>FCI</i>	105.76 *	-22.67	0.26
<i>Boundary</i>	ln(<i>Indirect/Direct</i>)	108.03 *	-5.75	0.33
<i>Boundary</i>	<i>IFI</i>	132.44 *	-47.43	0.11
<i>Boundary</i>	ln(<i>AGG</i>)	116.34 *	-9.66	0.14
<i>Boundary</i>	<i>HMG</i>	140.14 *	-14.09 *	0.04 *
<i>Boundary</i>	<i>AMP</i>	99.95 *	-14.55	0.40
<i>FCI</i>	ln(<i>Indirect/Direct</i>)	-0.16 *	0.28 *	0.00 *
<i>FCI</i>	<i>IFI</i>	-0.68 *	1.43 *	0.00 *
<i>FCI</i>	ln(<i>AGG</i>)	-0.24 *	0.31 *	0.00 *
<i>FCI</i>	<i>HMG</i>	0.17	0.07 *	0.05
<i>FCI</i>	<i>AMP</i>	0.22 *	0.81 *	0.00 *
ln(<i>Indirect/Direct</i>)	<i>IFI</i>	-1.74 *	4.92 *	0.00 *
ln(<i>Indirect/Direct</i>)	ln(<i>AGG</i>)	-0.21 *	1.07 *	0.00 *
ln(<i>Indirect/Direct</i>)	<i>HMG</i>	0.64	0.43 *	0.00 *
ln(<i>Indirect/Direct</i>)	<i>AMP</i>	1.36 *	2.73 *	0.00 *
<i>IFI</i>	ln(<i>AGG</i>)	0.31 *	0.22 *	0.00 *
<i>IFI</i>	<i>HMG</i>	0.47 *	0.09 *	0.00 *
<i>IFI</i>	<i>AMP</i>	0.64 *	0.52 *	0.00 *
ln(<i>AGG</i>)	<i>HMG</i>	0.91 *	0.36 *	0.00 *
ln(<i>AGG</i>)	<i>AMP</i>	1.51 *	2.37 *	0.00 *
<i>HMG</i>	<i>AMP</i>	2.98 *	0.45	0.08

* indicates significant at $\alpha = 0.05$

Table 5
Principle components factor analysis.

Variable	Factor 1	Factor 2	Uniqueness
<i>ln(TST)</i>	0.84	0.53	0.01
<i>ln(TSS)</i>	0.61	0.28	0.55
<i>Boundary</i>	-0.09	0.99	0.01
<i>FCI</i>	0.95	-0.04	0.09
<i>ln(Indirect/Direct)</i>	0.96	-0.02	0.07
<i>IFI</i>	0.98	-0.09	0.04
<i>ln(AGG)</i>	1.00	-0.07	0.01
<i>HMG</i>	0.34	-0.18	0.85
<i>AMP</i>	0.91	-0.01	0.17
>Loading Sums of Squares	5.81	1.40	
Proportion of Variance	0.65	0.16	
Cumulative Variance	0.65	0.80	

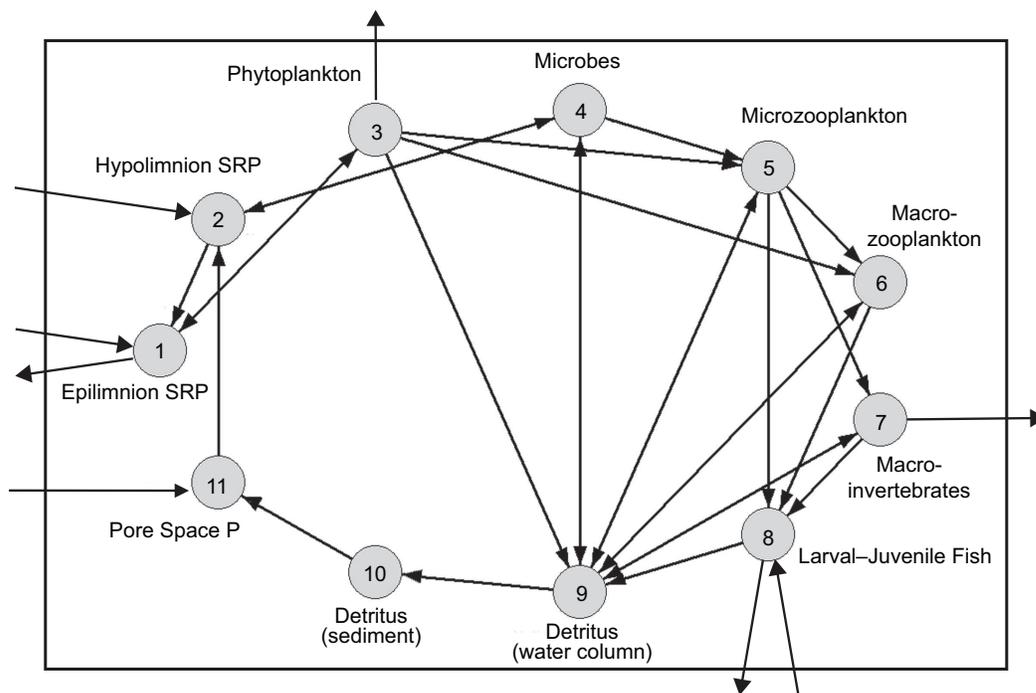


Fig. 1. Eleven compartment model of phosphorus flux in Lake Sidney Lanier, Georgia, USA.

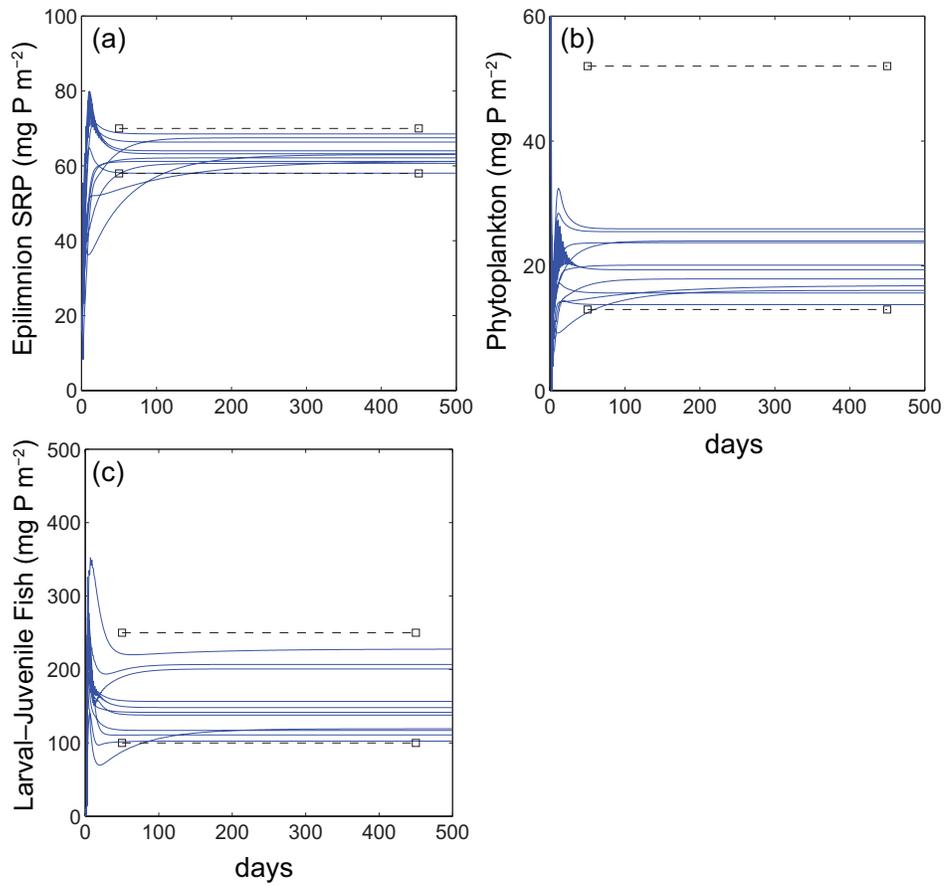


Fig. 2. Example of eleven successful model outputs for (a) epilimnion soluble reactive phosphorus, (b) phytoplankton, and (c) larval-juvenile fish. Dashed lines and square markers represent the constraints for the behavior definitions.

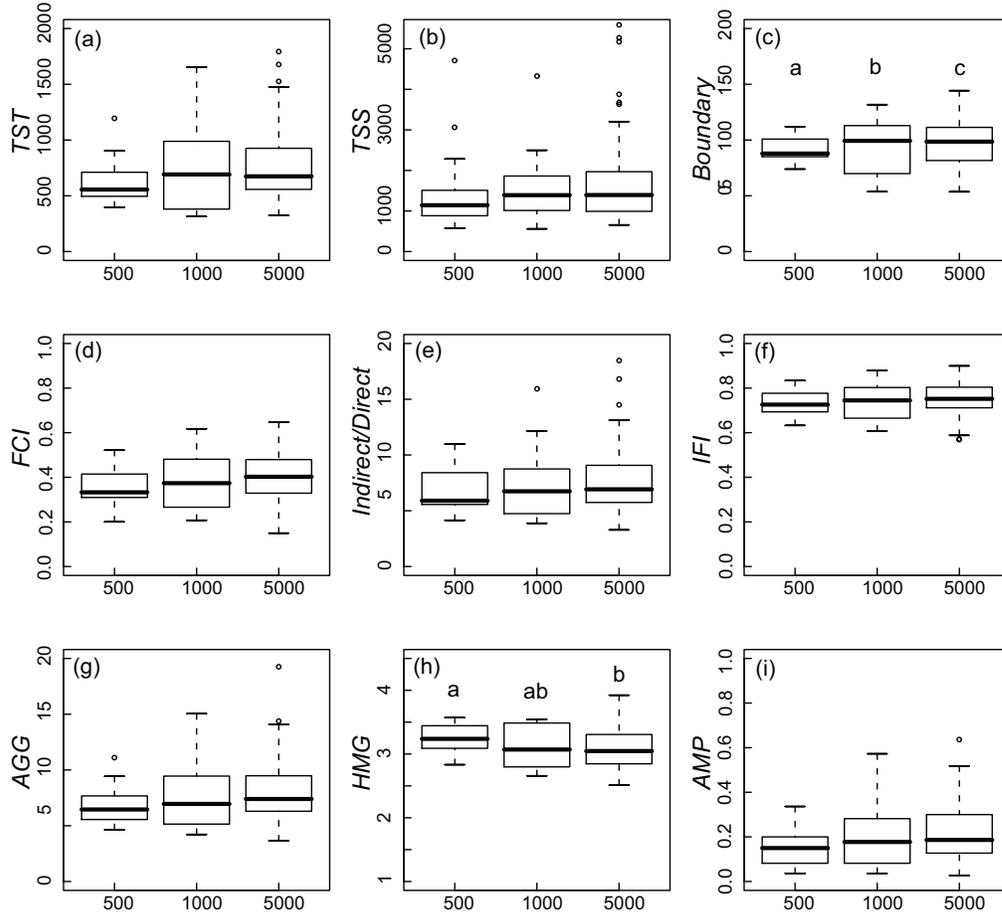


Fig. 3. Sample distributions of nine Network Environ Analysis indicators in plausible parameterizations of the Lake Lanier phosphorus model from simulations with 500, 1000, and 5000 trials. The trials generated 14, 18, and 90 plausible model parameterizations respectively. The indicators are (a) Total System Throughflow (TST), (b) Total System Storage (TSS), (c) Total Boundary Input ($Boundary$), (d) Finn cycling index (FCI), (e) ratio of indirect-to-direct flows ($Indirect/Direct$), (f) indirect flow index (IFI), (g) network aggradation (AGG), (h) network homogenization (HMG), and (i) network amplification (AMP).

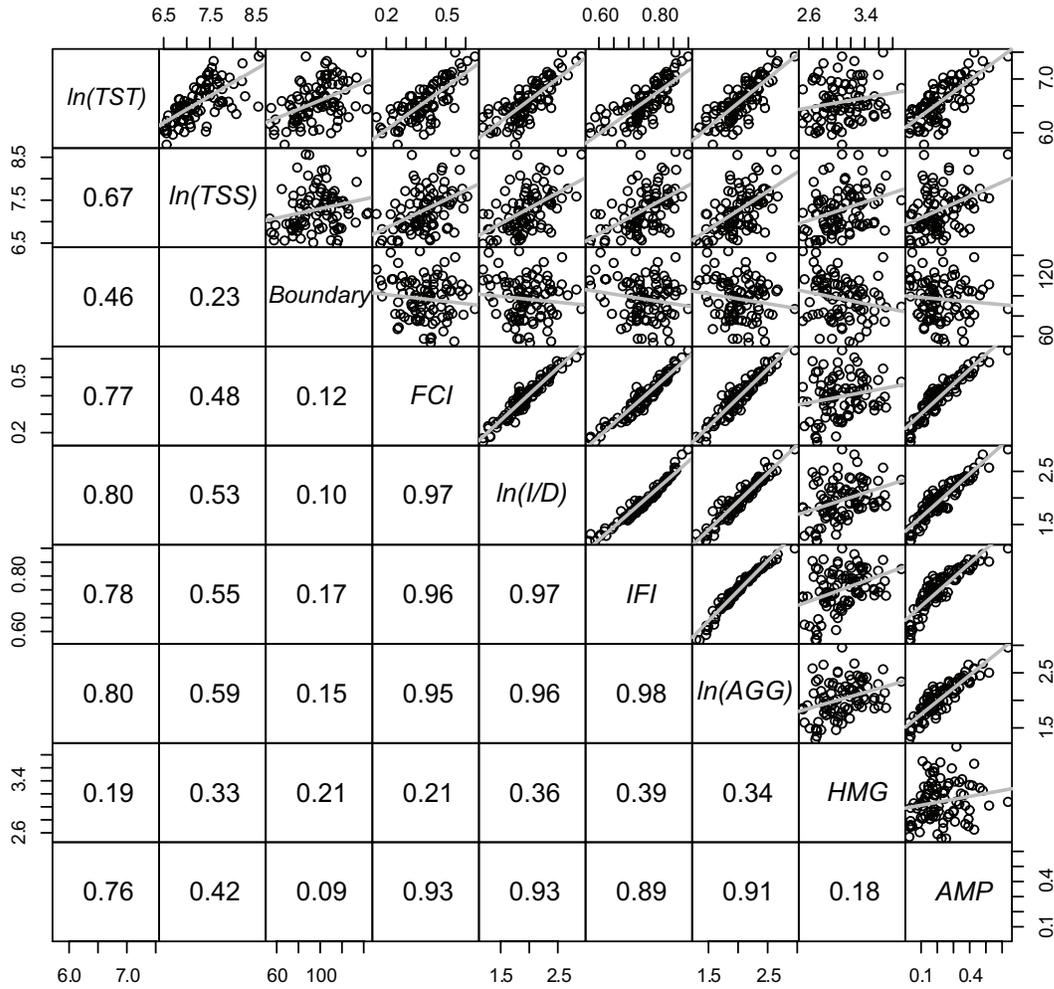


Fig. 4. Pairwise scatter plots of nine Network Environ Analysis indicators of ecosystem organization in 90 plausible parameterizations of the Lake Lanier phosphorus model. Scatter plots and ordinary least-squares regression lines are plotted above the principle diagonal and corresponding Pearson correlation coefficients are positioned below. Notice that we transformed TST , TSS , $Indirect/Direct$ (symbolized here as I/D), and AGG by the natural logarithm.

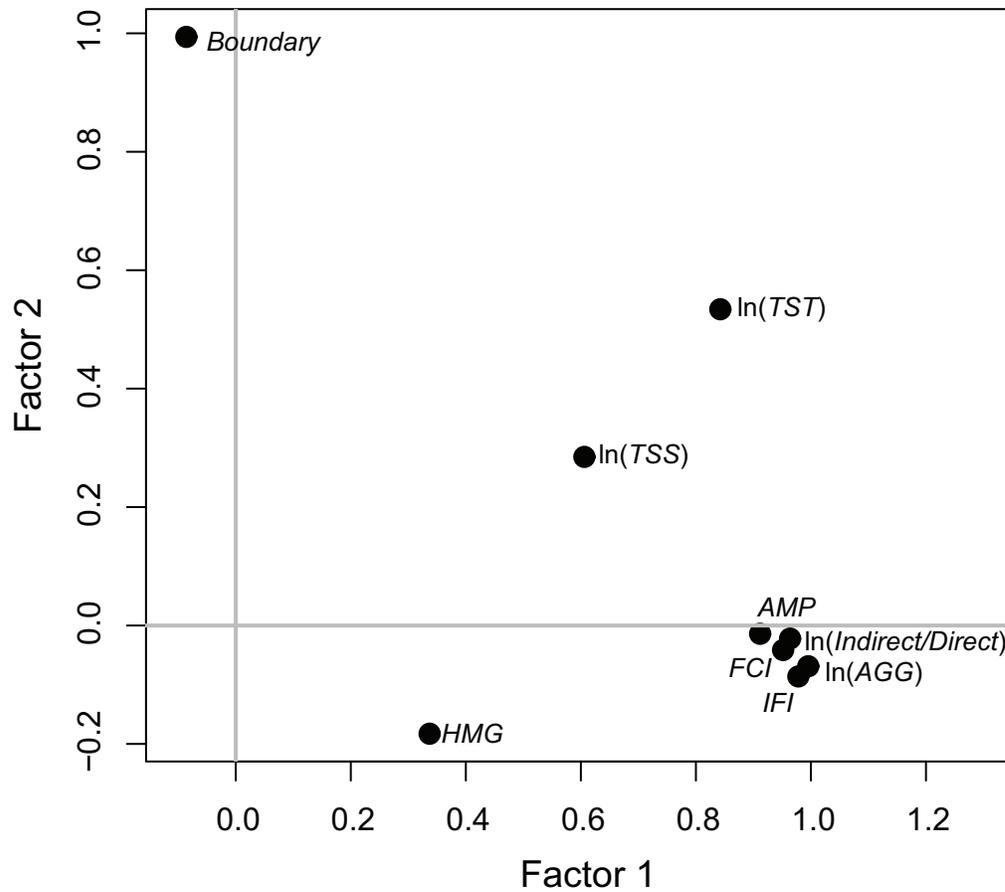


Fig. 5. Factor analysis loadings plot. Distance from the origin to an indicator implies the strength of its association with a factor. For example, *Boundary* is highly associated with Factor 2 and has little common variation with Factor 1.